

Package: PRIMEplus (via r-universe)

November 6, 2024

Title Study Design for Immunotherapy Clinical Trials

Version 1.0.16

Date 2024-01-09

Description Perform sample size, power calculation and subsequent analysis for Immuno-oncology (IO) trials composed of responders and non-responders.

Maintainer Bill Wheeler <wheelerb@imsweb.com>

Depends R (>= 3.5), survival, msm

License GPL-2

NeedsCompilation yes

Author Zhenzhen Xu [aut], Yongsoek Park [aut], Zhu Bin [aut], Bill Wheeler [cre]

Date/Publication 2024-01-10 14:03:13 UTC

Repository <https://wheelerb.r-universe.dev>

RemoteUrl <https://github.com/cran/PRIMEplus>

RemoteRef HEAD

RemoteSha 6ea68294404ed2446ca40ab64350bfbe0b6accc9

Contents

PRIMEplus-package	2
data	2
generate_data	3
getHazard	4
PRIMEplus.EM	5
PRIMEplus.LRT	6
PRIMEplus.Power	8
PRIMEplus.ReRand.LRT	9
PRIMEplus.SampleSize	11

Index	13
--------------	-----------

PRIMEplus-package

Study design for immunotherapy clinical trials

Description

Perform sample size, power calculation and subsequent analysis for Immuno-oncology (IO) trials composed of responders and nonresponders.

Details

This package is an extension of the Immunotherapy.Design R package but allows for response categories of more than two categories among the treatment population, such as complete responders, partial responders, as well as non-responders.

Author(s)

Zhenzhen Xu <Zhenzhen.Xu@fda.hhs.gov>, Yongsoek Park <yongpark@pitt.edu> and Bin Zhu <bin.zhu@nih.gov>

References

Xu, Z., Park, Y., Liu, K. and Zhu, B. Treating non-responders: pitfalls and implications for cancer immunotherapy trial design. *Journal of Hematology & Oncology* 13, 20 (2020).

Xu, Z., Zhu, B. and Park, Y. Designing immuno-oncology clinical trials composed of responders and nonresponders. *Statistics in Medicine*. (Under Revision).

data

Data for examples

Description

Data for examples.

Details

A data frame used in the examples.

Examples

```
data(data, package="PRIMEplus")
```

```
# Display some of the data  
data[1:5, ]
```

generate_data	<i>Simulated data</i>
---------------	-----------------------

Description

Generate simulated data

Usage

```
generate_data(N=500, rand_ratio=0.5, effect_p=0.6, enroll_rate=380*0.25/6,
              lambda1=0.117, HR=0.5, tau=12*5, t1=1)
```

Arguments

N	Maximum sample size
rand_ratio	Allocation ratio
effect_p	Vector for proportion of responders in the treatment arm at baseline (see details)
enroll_rate	Enrollment rate in subjects per month
lambda1	Baseline hazard in terms of months
HR	Vector of hazard ratios for responders against controls (see details)
tau	Total study duration
t1	Delayed duration

Details

The length and order of `effect_p` must be the same as `HR`. Both of these vectors should contain information only for the groups of responders. For example, if there are full responders and partial responders, then `effect_p` and `HR` would be vectors of length two.

Value

A data frame with columns:

Name	Description
id	id variable
trt	treatment allocation: 1 = treatment arm
D1	patient's response status for group 1
D2	patient's response status for group 2
Dm	patient's response status for non-responders
tau	total study duration
enroll_time	patients' enrollment times
time_to_event	patients' event times
event_status	censoring indicator
X	observational time
t1	delayed duration

Author(s)

Zhenzhen Xu <Zhenzhen.Xu@fda.hhs.gov>, Yongsoek Park <yongpark@pitt.edu> and Bin Zhu <bin.zhu@nih.gov>

Examples

```
data <- generate_data()
data[1:5, ]
```

getHazard	<i>Compute initial estimates for the baseline hazard</i>
-----------	--

Description

Calls the coxph function to compute initial estimates for the baseline hazard

Usage

```
getHazard(time, treatment, event_status, t.fail.o=NULL)
```

Arguments

time	Vector of times.
treatment	Binary vector of treatments (1=subject received treatment).
event_status	Binary vector of event status (1=subject experienced an event).
t.fail.o	NULL or vector of event times.

Value

List containing the baseline hazards ordered by the event times.

Author(s)

Zhenzhen Xu <Zhenzhen.Xu@fda.hhs.gov> and Bin Zhu <bin.zhu@nih.gov>

See Also

[PRIMEplus.EM](#)

Examples

```
data(data, package="PRIMEplus")
lambda0 <- getHazard(data[, "X"], data[, "trt"], data[, "event_status"])$hazard
lambda0[1:10]
```

PRIMEplus.EM

*EM algorithm***Description**

EM algorithm

Usage

```
PRIMEplus.EM(data, effect_p, beta0, time.var="X", trt.var="trt",
             status.var="event_status", id.var="id", t1=1, lambda0=NULL,
             stopTol=1e-4, maxiter=100000, print=0)
```

Arguments

<code>data</code>	Data frame or matrix containing a time-to-event variable (<code>time.var</code>), a treatment variable (<code>trt.var</code>), and a censoring variable (<code>status.var</code>).
<code>effect_p</code>	Vector of proportions for groups of responders in the treatment arm at baseline (see details).
<code>beta0</code>	Vector or matrix of initial estimates for the log-hazard ratios (see details).
<code>time.var</code>	Time-to-event variable name in data. The default is "X".
<code>trt.var</code>	Binary treatment variable name in data coded as 0 for controls and 1 for subjects that received treatment.
<code>status.var</code>	Name of the binary censoring variable in data coded as 0 for censored subjects and 1 for subjects that experienced an event.
<code>id.var</code>	NULL or subject id variable in data. The default is "id".
<code>t1</code>	Delayed duration. The default is 1.
<code>lambda0</code>	NULL or vector of initial estimates for the baseline hazards corresponding to the ordered event times.
<code>stopTol</code>	Stopping tolerance. The default is 1e-4.
<code>maxiter</code>	Maximum number of iterations. The default is 100000.
<code>print</code>	0-2 to print information. Larger values will print more information. The default is 0.

Details

The EM algorithm is sensitive to the initial values of the log-hazard ratios (`beta0`), so different initial estimates should be tried to ensure the maximum log-likelihood is obtained. Thus, `beta0` can be a vector or matrix, where in the case of a matrix, each row corresponds to a different set of initial estimates. Each set of initial estimates must contain distinct non-zero values. The length and order of `effect_p` must be the same as the columns of `beta0`. Both of these should contain information only for the groups of responders. For example, if there are full responders and partial responders, then `effect_p` would be a vector of length two, and `beta0` would be a vector of length two or a matrix with two columns.

Value

A list containing the objects:

Name	Description
converged	TRUE if EM algorithm converged
beta	final log(hazard ratio) estimates of responders versus controls
lambda	final estimates of baseline hazards
probResponder	estimated probability of a subject being a responder
loglike	log-likelihood value at the final estimates
loglike.marg	marginal log-likelihood value at the final estimates

Author(s)

Zhenzhen Xu <Zhenzhen.Xu@fda.hhs.gov>, Yongsoek Park <yongpark@pitt.edu> and Bin Zhu <bin.zhu@nih.gov>

See Also

[getHazard](#), [PRIMEplus.LRT](#)

Examples

```
data(data, package="PRIMEplus")
effp <- c(0.3, 0.3)
beta0 <- c(log(0.4), log(0.6))
ret <- PRIMEplus.EM(data, effp, beta0)
ret$beta
```

PRIMEplus.LRT

Likelihood Ratio Test

Description

PRIMEplus likelihood ratio test

Usage

```
PRIMEplus.LRT(data, effect_p, beta0, time.var="X", trt.var="trt",
  status.var="event_status", id.var="id", t1=1, lambda0=NULL,
  stopTol=1e-4, maxiter=100000, print=0)
```

Arguments

data	Data frame or matrix containing a time-to-event variable (<code>time.var</code>), a treatment variable (<code>trt.var</code>), and a censoring variable (<code>status.var</code>).
effect_p	Vector of proportions for groups of responders in the treatment arm at baseline (see details).

beta0	Vector or matrix of initial estimates for the log-hazard ratios (see details).
time.var	Time-to-event variable name in data. The default is "X".
trt.var	Binary treatment variable name in data coded as 0 for controls and 1 for subjects that received treatment.
status.var	Name of the binary censoring variable in data coded as 0 for censored subjects and 1 for subjects that experienced an event.
id.var	NULL or subject id variable in data. The default is "id".
t1	Delayed duration. The default is 1.
lambda0	NULL or vector of initial estimates for the baseline hazards corresponding to the ordered event times.
stopTol	Stopping tolerance. The default is 1e-4.
maxiter	Maximum number of iterations. The default is 100000.
print	0-2 to print information. Larger values will print more information. The default is 0.

Details

The EM algorithm is sensitive to the initial values of the log-hazard ratios (β_0), so different initial estimates should be tried to ensure the maximum log-likelihood is obtained. Thus, β_0 can be a vector or matrix, where in the case of a matrix, each row corresponds to a different set of initial estimates. Each set of initial estimates must contain distinct non-zero values. The length and order of `effect_p` must be the same as the columns of β_0 . Both of these should contain information only for the groups of responders. For example, if there are full responders and partial responders, then `effect_p` would be a vector of length two, and β_0 would be a vector of length two or a matrix with two columns.

Value

A list containing the objects:

Name	Description
converged	TRUE if EM algorithm converged
beta	final log(hazard ratio) estimates of responders versus controls
lambda	final estimates of baseline hazards
probResponder	estimated probability of a subject being a responder
loglike	log-likelihood value at the final estimates
loglike.marg	marginal log-likelihood value at the final estimates
loglike.marg.0	marginal log-likelihood value under the null hypothesis
LRT	likelihood-ratio test statistic
pvalue	p-value of the likelihood ratio test

Author(s)

Zhenzhen Xu <Zhenzhen.Xu@fda.hhs.gov>, Yongsoek Park <yongpark@pitt.edu> and Bin Zhu <bin.zhu@nih.gov>

See Also[PRIMEplus.EM](#)**Examples**

```

data(data, package="PRIMEplus")
effp <- c(0.3, 0.3)
beta0 <- c(log(0.4), log(0.6))
ret <- PRIMEplus.LRT(data, effp, beta0)
ret$LRT
ret$pvalue

```

PRIMEplus.Power	<i>Power</i>
-----------------	--------------

Description

Compute the power using LRT Re-randomization test

Usage

```

PRIMEplus.Power(nmax=500, rand_ratio=0.5, effect_p=0.6,
  enroll_rate=380*0.25/6, lambda1=0.117, HR=0.5, tau=12*5, t1=1,
  maxiter=100000, stopTol=1e-4, alpha=0.05, num_rand=1000, nsim=10000,
  print=0, min.sample.size=50, min.n.event=5, min.per.trt=0.25)

```

Arguments

nmax	Sample size
rand_ratio	Probability of assignment to treatment arm
effect_p	Vector for proportion of responders in the treatment arm at baseline (see details)
enroll_rate	Enrollment rate in subjects per month
lambda1	Baseline hazard in terms of months
HR	Vector of hazard ratios for responders against controls (see details)
tau	Total study duration
t1	Delayed duration in months
maxiter	Maximum number of iterations in the EM algorithm. The default is 100000.
stopTol	Stopping tolerance in the EM algorithm. The default is 1e-4.
alpha	Significance level. The default is 0.05.
num_rand	The number of replications in the re-randomization test. The default is 1000.
nsim	The number of simulations. The default is 1000.
print	0 or 1 to print information. The default is 0.
min.sample.size	Minimum sample size. The default is 50.
min.n.event	Minimum number of events. The default is 5.
min.per.trt	Minimum proportion of controls and treated subjects. The default is 0.25.

Details

The length and order of `effect_p` must be the same as `HR`. Both of these vectors should contain information only for the groups of responders. For example, if there are full responders and partial responders, then `effect_p` and `HR` would be vectors of length two.

For each simulation, a simulated data set is created from the `generate_data` function and then an estimated p-value is computed by calling `PRIMEplus.ReRand.LRT`. The power is calculated as the proportion of iterations whose estimated p-value was less than or equal to `alpha`.

Value

A list containing the power and the number of simulated datasets used in the calculation.

Author(s)

Zhenzhen Xu <Zhenzhen.Xu@fda.hhs.gov>, Yongsoek Park <yongpark@pitt.edu> and Bin Zhu <bin.zhu@nih.gov>

See Also

[PRIMEplus.ReRand.LRT](#)

PRIMEplus.ReRand.LRT *Randomization Test*

Description

PRIMEplus randomization likelihood ratio test

Usage

```
PRIMEplus.ReRand.LRT(data, effect_p, beta0, time.var="X", trt.var="trt",
  status.var="event_status", id.var="id", t1=1, lambda0=NULL,
  stopTol=1e-4, maxiter=100000, print=0, num_rand=1000)
```

Arguments

<code>data</code>	Data frame or matrix containing a time-to-event variable (<code>time.var</code>), a treatment variable (<code>trt.var</code>), and a censoring variable (<code>status.var</code>).
<code>effect_p</code>	Vector of proportions for groups of responders in the treatment arm at baseline (see details).
<code>beta0</code>	Vector or matrix of initial estimates for the log-hazard ratios (see details).
<code>time.var</code>	Time-to-event variable name in data. The default is "X".
<code>trt.var</code>	Binary treatment variable name in data coded as 0 for controls and 1 for subjects that received treatment.
<code>status.var</code>	Name of the binary censoring variable in data coded as 0 for censored subjects and 1 for subjects that experienced an event.

id.var	NULL or subject id variable in data. The default is "id".
t1	Delayed duration. The default is 1.
lambda0	NULL or vector of initial estimates for the baseline hazards corresponding to the ordered event times.
stopTol	Stopping tolerance. The default is 1e-4.
maxiter	Maximum number of iterations. The default is 100000.
print	0-2 to print information. Larger values will print more information. The default is 0.
num_rand	The number of randomizations. The default is 1000.

Details

The EM algorithm is sensitive to the initial values of the log-hazard ratios (β_0), so different initial estimates should be tried to ensure the maximum log-likelihood is obtained. Thus, β_0 can be a vector or matrix, where in the case of a matrix, each row corresponds to a different set of initial estimates. Each set of initial estimates must contain distinct non-zero values. The length and order of `effect_p` must be the same as the columns of β_0 . Both of these should contain information only for the groups of responders. For example, if there are full responders and partial responders, then `effect_p` would be a vector of length two, and β_0 would be a vector of length two or a matrix with two columns. The initial values are only used for the observed data; each randomization uses the MLE estimates as initial estimates.

Value

A list containing the objects:

Name	Description
<code>pvalue.LRT</code>	p-value of the randomization test based on the likelihood ratio test
<code>pvalue.loglike.marg</code>	p-value of the randomization test based on the marginal likelihood
<code>n.randomizations</code>	the number of randomizations that the p-values are based on

Author(s)

Zhenzhen Xu <Zhenzhen.Xu@fda.hhs.gov>, Yongsoek Park <yongpark@pitt.edu> and Bin Zhu <bin.zhu@nih.gov>

See Also

[PRIMEplus.LRT](#)

Examples

```
data(data, package="PRIMEplus")
effp <- c(0.3, 0.3)
beta0 <- c(log(0.4), log(0.6))
ret <- PRIMEplus.ReRand.LRT(data, effp, beta0, num_rand=100)
ret
```

 PRIMEplus.SampleSize *Sample Size*

Description

Compute the sample size for a given power

Usage

```
PRIMEplus.SampleSize(power=0.8, rand_ratio=0.5, effect_p=0.6, enroll_rate=380*0.25/6,
  lambda1=0.117, HR=0.5, tau=12*5, t1=1, maxiter=100000, stopTol=1e-4,
  alpha=0.05, num_rand=1000, nsim=10000, min.N=100, max.N=700,
  tol.power=0.01, tol.N=1, print=1,
  min.sample.size=50, min.n.event=5, min.per.trt=0.25)
```

Arguments

power	The desired power. The default is 0.8.
rand_ratio	Allocation ratio
effect_p	Vector for proportion of responders in the treatment arm at baseline (see details)
enroll_rate	Enrollment rate in subjects per month
lambda1	Baseline hazard in terms of months
HR	Vector of hazard ratios for responders against controls (see details)
tau	Total study duration
t1	Delayed duration
maxiter	Maximum number of iterations in the EM algorithm. The default is 100000.
stopTol	Stopping tolerance in the EM algorithm. The default is 1e-4.
alpha	Significance level. The default is 0.05.
num_rand	Number of replications in the re-randomization test. The default is 1000.
nsim	Number of simulations in computing power (see Details). The default is 10000.
min.N	Lower bound for the sample size. The default is 100.
max.N	Upper bound for the sample size. The default is 700.
tol.power	Stopping tolerance for the power. The default is 0.01.
tol.N	Stopping tolerance for the sample size. The default is 1.
print	0 or 1 to print information. The default is 1.
min.sample.size	Minimum sample size. The default is 50.
min.n.event	Minimum number of events. The default is 5.
min.per.trt	Minimum proportion of controls and treated subjects. The default is 0.25.

Details

The length and order of `effect_p` must be the same as `HR`. Both of these vectors should contain information only for the groups of responders. For example, if there are full responders and partial responders, then `effect_p` and `HR` would be vectors of length two.

This uses a bisection method to estimate the sample size. At each iteration, the estimated power `power_est` is computed using [PRIMEplus.Power](#) for a given sample size holding all other parameters fixed. The algorithm terminates when $\text{abs}(\text{power} - \text{power_est}) \leq \text{tol} \cdot \text{power}$ or when the length of the estimated interval containing the sample size is less than or equal to $\text{tol} \cdot N$.

NOTE:

It is important to note that the power for a given sample size is estimated by running a simulation. Thus, by setting a different seed, a different result may be returned. Therefore, to ensure a more precise estimated sample size, set the option `nsim` to a large value and/or run this function several times by setting different seeds and examine the distribution of returned sample sizes.

Value

A list containing the sample size and power.

Author(s)

Zhenzhen Xu <Zhenzhen.Xu@fda.hhs.gov>, Yongsoek Park <yongpark@pitt.edu> and Bin Zhu <bin.zhu@nih.gov>

See Also

[PRIMEplus.Power](#)

Index

- * **EM**
 - PRIMEplus.EM, 5
- * **LRT**
 - PRIMEplus.LRT, 6
- * **data**
 - data, 2
- * **hazard**
 - getHazard, 4
- * **package**
 - PRIMEplus-package, 2
- * **permutation**
 - PRIMEplus.ReRand.LRT, 9
- * **power**
 - PRIMEplus.Power, 8
- * **sample size**
 - PRIMEplus.SampleSize, 11
- * **simulation**
 - generate_data, 3

data, 2

generate_data, 3, 9

getHazard, 4, 6

PRIMEplus (PRIMEplus-package), 2

PRIMEplus-package, 2

PRIMEplus.EM, 4, 5, 8

PRIMEplus.LRT, 6, 6, 10

PRIMEplus.Power, 8, 12

PRIMEplus.ReRand.LRT, 9, 9

PRIMEplus.SampleSize, 11